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The Queen's University of Belfast
School of Biology and Biochemistry
BELFAST
BT9 7BL
Northern Ireland

Patents ADP number *(if you know it)*

If the applicant is a corporate body, give the country/state of its incorporation — UK 7422330001

4. Title of the invention

"Medicament"

5. Name of your agent *(if you have one)*

"Address for service" in the United Kingdom to which all correspondence should be sent *(including the postcode)*

Murgitroyd & Company
373 Scotland Street
GLASGOW
G5 8QA
United Kingdom

Patents ADP number *(if you know it)* — 1198013

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"Medicament"

This invention relates to the use of modified peptides for the treatment of angiogenic diseases and inhibition of the secondary spread of cancer, as well as treatment of *Candida spp.* infections.

Angiogenic diseases are those disorders which are directly caused by, or complicated by the inappropriate growth of new blood vessels. The major angiogenic diseases include the common metastatic solid tissue cancers (breast, gastrointestinal, lung, prostatic, etc), diabetic retinopathy, neovascular glaucoma, rheumatoid arthritis and psoriasis. Angiogenesis is the rate-limiting step in the growth of secondary tumours; inhibition of their neovascularisation is known to stop their growth.

In this field it is already known that the native ligand of the 67kDa laminin receptor (67LR) is encompassed by the linear sequence of amino acids 925-933 of the laminin b1 chain (numbering refers to the mature murine laminin b1). Synthetic laminin $b1_{925-933}$ (single letter amino acid code: CDPGYIGSR-NH_2) has been shown to inhibit tumour establishment in mice, by

inhibiting attachment of tumour cells to basement membranes. It has also ben demonstrated that laminin $b1_{925-933}$ inhibits angiogenesis in the chick.

However, synthetic laminin-derived peptide (laminin $b1_{925-933}$) stimulates angiogenic events in mammalian cells (in which it acts as a pure 67LR agonist), making it useless as the basis of a human therapy. This dual action suggests that the effects of EGF on motility are mediated by downstream action of 67LR.

It is one object of the present invention to provide a medicament to treat angiogenic diseases.

The present invention provides a peptide factor derived from murine epidermal growth factor (EFD) peptide for use in the preparation of a medicament for the treatment of angiogenic diseases.

The mechanism by which EGF derived peptides inhibit new blood vessel formation is through their antagonism of the high affinity 67 kDa laminin receptor (67LR) found on endothelial cells.

The peptides have the additional effect of inhibiting tumour cell attachment to basement membranes, and may be used to prevent solid cancer spread in cases where cancer cells have been identified circulating in the blood.

The modified peptides are protected from proteolytic degradation by substitution of key residues with unnatural amino acid analogues at susceptible bonds, such as tyrosine analogues (at position 5) and arginine analogues (at position 9). The peptides are capped at *N*- and *C*-termini (with acetyl and amide groups

respectively) and at the thiol groups of the cysteines (with acetamido methyl groups).

Typically the peptide is an antagonist of the 67kDa Laminin Receptor (67LR).

The peptide factor is based on amino acid residues 32 to 42 of murine epidermal growth factor (mEGF).

The amino acid sequence of mEGF- (33-42) is CVIGYSGDRC.

Preferably the sequence of peptide factor is modified from the natural sequence to protect the peptides from protease attack.

Preferred substitutions include the use of tyrosine analogues at position 5 and arginine analogues at position 9.

Preferably the peptide factor is capped at the N terminal with an acetyl group.

Preferably the peptide factor is capped at the C terminal with an amide group.

Preferably the thiol groups of cysterines are capped with acetamido methyl groups.

In one embodiment the synthetic peptide has the sequence

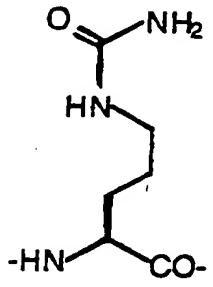
Acetyl-C-[S-Acm]-VIGYSGDR-C-[S-Acm]NH_2

A preferred tyrosine analogue is Tic-OH.

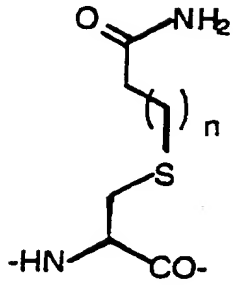
A preferred arginine analogue is Citrulline.

The structure of Citrulline and other potential arginine analogues are shown below.

Citrulline and analogues



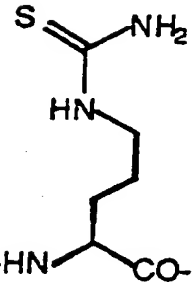
citrulline



{n = 1,2}

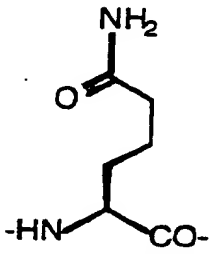
cysteine-derived analogues

{prepared by reaction of cysteine with $Br-(CH_2)_n-CONH_2$}



thiono-citrulline

{prepared by reaction of ornithine with ammonium isothiocyanate}



homo-glutamine

Preferably the peptide is truncated to a shorter peptide without losing its antagonistic character.

The invention further provides a peptide agonist.

The agonist (typically containing Tic-OH substitution for tyrosine) might be useful in healing endiothelial

cell wounding.

For example, corneal endothielial cells can be damaged during cataract operations and this damage does not self-repair because these endothelial cells do not divide. Healing can only be by cell migration and spreading, and this may be promoted by the agonist.

In order to explore possible conformations for the parent $mEGF_{33-42}$ peptide, it was modelled using molecular dynamics. Based on these conformations we were able to predict a strategy to provide proteolytic protection by being able to identify residues that were important to the maintenance of a three-dimensional conformation essential for 67LR recognition.

The following is a description of some examples of modifications and uses of the invention.

1. On the basis of the modelled structures, we found that the arginine residue participated in H-bonding, and speculated that this charge may not be important. We synthesised a peptide based on $mEGF_{33-42}$, in which the arginine residue at position 41 was replaced by citrulline (an uncharged arginine mimetic with similar H-bonding potential). This peptide provided to act as a more potent 67LR antagonist and was found to be resistant to trypsin degradation.

2. Double substitution of $tyrosine_{37}$ with Tic-OH and $Arginine_{41}$ with citrulline, to produce a $mEGF_{33-42}$-derived peptide resistant to both chymotrypsin-like and trypsin-like proteases.

3. Replacement of susceptible peptide bonds in $mEGF_{33-}$

$_{42}$ with protease-resistant peptide bond isosteres (such as thionopeptide or methylene amino bonds).

4. Conformationally restricted analogues may give improved potency due to the essential 3-dimensional conformation being stabilised. For example, it should be possible to increase the rigidity of the molecule by replacing each of the central glycine residues in turn by α,α-dialkyl substituted amino acids such as α-amino isobutyric acid (AIB) or aminocyclopropane carboxylic acid (ACPCA). Alternatively, the helical turn (which we have identified as essential) could be stabilised by bridging with suitable intra-chain linkers, such as a disulphide bond between *N*- and *C*-terminal [*D*] or [*L*]-cysteines.

Alternative Uses

1. Some microbial pathogens such as *Candida albicans*, express 67LR and use this as a means of attaching to human basement membranes. It is conceivable that such infections could be abolished by treatment with $mEGF_{33-42}$-derived peptides, which would prevent the microbes from adhering to the host.

Advantages

The advantages of the invention, and the ways in which disadvantages of previously known arrangements are overcome include:

1. Unlike the native 67LR ligand (laminin $b1_{925-933}$), which is angiogenic in human models, the $mEGF_{33-42}$-derived agents are anti-angiogenic in human

models.

2. $mEGF_{33-42}$ has the advantage of inhibiting both laminin- and EGF-stimulated angiogenesis.

3. $mEGF_{33-42}$ prevents tumour cell attachment to basement membranes.

Examples

The purpose of the investigation was to determine the molecular target of $mEGF_{(33-42)}$ and to identify the amino acids that are essential for receptor recognition. In addition, the key residues which confer laminin antagonism on $mEGF_{(33-42)}$ were examined.

Two lead compounds were invesigated; synthetic laminin B1 sequence CDPGYIGSR-NH_2 and $mEGF_{(33-42)}$ sequence AcC(Acm)-VIGYSGDRC-(Acm)-NH_2. Bearing in mind the pure antagonism of the murine EGF peptide, the aims of this study were to identify the key residues responsible for these contrasting activities using alanine scanning, in the context of developing anti-angiogenic drugs for retinopathy treatment.

In addition, using residue exchange between the two peptides and molecular modelling to predict three-dimensional structure, we wished to further investigate the role of individual $mEGF_{(33-42)}$ residues in laminin antagonism. A logical series of peptides was synthesised and screened for receptor interaction, cell adhesion and motility properties (Table 1a and 1b).

MATERIALS AND METHODS

Peptide synthesis

Peptide sequences based on and $mEGF_{(33-42)}$ were synthesised on a model 432A peptide synthesiser (Applied Biosystems, Warrington, UK), using standard solid-phase Fmoc procedure (Fields 1990). Synthesis of the peptides required successive additions of derivatized amino acids to form a linear product.

Peptides were purified after synthesis using reverse phase HPLC and purity confirmed by automated amino acid analysis and electrospray mass spectrometry. All peptide sequences were stored in the presence of desiccant at -20°C until required for biological assay.

Laminin receptor antibody production

a. Preparation of MAPs

The peptide sequence (PTEDWSAQPATEDWSAAPTA), corresponding to the COOH-terminal end of the human laminin receptor, was used as the antigen template. Derivation of the peptide, based on a CN-Br cleavage fragment of the cDNA sequence encoding human laminin receptor, has been described elsewhere (Wewer et al 1986). The antigen was synthesised as an octomeric peptide derivative (MAPs) using automated Fmoc procedure (Tam 1988).

$mEGF_{(33-42)}$	acetyl	ACM Cys	Val	Ile	Gly	Tyr	Ser	Gly	Asp	Arg	ACM Cys-NH_2
I	acetyl	ACM Cys	Val	Ile	Gly	Tyr	Ile	Gly	Asp	Arg	ACM Cys-NH_2
II	acetyl	ACM Cys	Val	Ile	Gly	Tyr	Ser	Gly	Ser	Arg	ACM Cys-NH_2
III	acetyl	ACM Cys	Val	Ile	Gly	Tyr	Ile	Gly	Ser	Arg	ACM Cys-NH_2
IV	acetyl	ACM Cys	Val	Ile	Gly	Tyr	Ser	Gly	Asp	Cit	ACM Cys-NH_2
V	acetyl	ACM Cys	Val	Ile	Gly	OH Tic	Ser	Gly	Asp	Arg	ACM Cys-NH_2

Table 1a : Peptide substi

Table 1b: Peptide substitution (alanine scanning)

$mEGF_{(33-42)}$	acetyl	ACM Cys	Val	Ile	Gly	Tyr	Ser	Gly	Asp	Arg	ACM Cys-NH_2
VI	acetyl	ACM Cys	Val	**Ala**	Gly	Tyr	Ser	Gly	Asp	Arg	ACM Cys-NH_2
VII	acetyl	ACM Cys	**Ala**	Ile	Gly	Tyr	Ser	Gly	Asp	Arg	ACM Cys-NH_2
VIII	acetyl	**Ala**	Val	Ile	Gly	Tyr	Ser	Gly	Asp	Arg	ACM Cys-NH_2
IX	acetyl	ACM Cys	Val	Ile	Gly	Tyr	Ser	Gly	Asp	Arg	**Ala-NH_2**
X	acetyl	ACM Cys	Val	Ile	Gly	Tyr	**Ala**	Gly	Asp	Arg	ACM Cys-NH_2

b. Immunisation schedule

A pre-immune test bleed (5ml) was obtained from the marginal ear vein of a male New Zealand White rabbit (3.2 kg). The bleed was allowed to clot for 2 h at room temperature after which its edge was detached from the wall of the collection vessel. The clot was then allowed to contract overnight at 4°C. Serum was then removed and the residual material pelleted out by centrifugation (10 min at 2,500 g). Extracted serum (3.5 ml) was then frozen at -20°C until required.

Immunogen was prepared by the emulsion of MAPs (0.5 g antigen in 0.5 ml PBS) in an equivalent volume of adjuvant (Alum Imject; Pierce, Chester, UK). The animals immune system was primed by introducing immunogen (50 µg) through subcutaneous injection at different sites on the animals back. The rabbit was boosted by both subcutaneous and intramuscular injection, 21 days after priming, using an increased dose of immunogen (800 µg). Subsequent boosts were performed by intramuscular injection after a further 14 days (800 µg immunogen), and thereafter at 21 day intervals.Test bleeds were taken 2 days after each boost and the serum extracted as described above. The animal was boosted and bled a total of three times.

c. Enzyme-linked immunoabsorbent assay

ELISA was used to determine the specificity of the antibody prepared against the synthetic MAPs peptide and to determine the efficacy of binding with respect to that of the linear precursor.

Peptides were dissolved in distilled water and diluted to 10 µg/ml in coating buffer. Aliquots (100 µl) of

either linear or MAPs peptide were then added to the wells of microtitre plates (Microtest III; Becton Dickinson Ltd., Oxford, UK) and incubated overnight at 37°C. The wells were then rinsed with 100 µl wash buffer and air dried. Excess adsorption sites were blocked (1 h incubation at 22°C) by the addition of 10% casein in PBS (0.1 ml/well). Subsequent to the removal of casein solution by aspiration, wells were again rinsed with wash buffer and air dried.

Antisera or pre-immune sera were then serially diluted in PBS and 100 µl of each incubated in peptide coated wells for 1 h at 37°C. After rinsing (0.1 ml wash buffer), 100 µl per well of 5 µg/ml secondary antibody (horse-radish peroxidase-conjugated goat anti-rabbit IgG; Amersham International, Aylesbury, UK) was added to each well and the plates incubated at 37°C for 1 h.

Wells were again rinsed with wash buffer and 0.1 ml substrate solution (TMB peroxidase) added to each. The plate was then incubated at 22°C for 30 min and the colour reaction stopped by the addition of 0.5M H_2SO_4 (0.1 ml/well). Absorbence was measured at 450 nm on a Titertek Multiscan plate reader.

d. Purification of IgG fraction

Anti-laminin receptor antiserum was purified using immobilised protein G-sepharose columns (Pharmacia Biotech, Uppasla, Sweden). The columns were equilibrated with 20 ml sodium phosphate buffer (pH 7.0). Antiserum was diluted 1:4 in the same buffer and a 1 ml aliquot loaded onto the column (flow rate 150 ml/h, fraction size 2.5 ml). After exclusion of the unbound fraction, as determined by absorbence at 280nm, the IgG component of the antiserum was eluted with 0.1M

glycine-HCl (pH 2.7), into tubes containing 0.1 ml Tris (1M), pH 9.0. The eluted IgG fractions were bulked and stored at -20°C until required.

Maintenance of cell cultures

Cancer and endothelial cells were maintained in either DMEM (T47-D) or RPMI (SK HEP-1) media, supplemented with 10% FCS, 100 IU/ml penicillin and 100 µg/ml streptomycin. Cells were incubated at 37°C in a humidified atmosphere of 95% air: 5% CO_2 and media refreshed as required. Cultures (at 80-85% confluence) were routinely passed on removal from monolayer by the action of trypsin (0.25%) and EDTA (0.02%) in CFS.

The viability of cell populations following trypsinisation was determined by the trypan blue vital dye exclusion test. Populations confirmed as being in excess of 95% viable were used in all studies.

Media were screened for possible bacterial or fungal contamination by incubating 1ml aliquots with both nutrient and Saboraud dextrose broths (Oxoid Ltd., Basingstoke, UK). Cell populations were routinely monitored for sub-clinical infections by periodically culturing in the absence of antibiotics.

Both cell lines and media were examined for the presence of contaminating *Mycoplasma* spp. by the method of Chen (1977).

Determination of cell numbers

Single cell suspensions were quantified using an automated counter (Coulter Electronics, Harpenden, UK). A 1 ml aliquot of cell suspension was diluted 1 in 20

in Isoton and 0.5 ml samples counted. The mean of 5 counts was taken and the total number of cells determined. Estimates of cell number were confirmed by counting in a haemocytometer.

For microtitre end-point assays, cell numbers were estimated from the crystal violet staining index of the cell line (Kanamaru and Yoshida 1989). Briefly, after removal of media from the assay system cells were fixed with formaldehyde (10% in PBS), and washed with distilled H_2O. Aliquots (100 µl) of crystal violet solution (0.1% in distilled H_2O) were added to each well and the plates allowed to stand for 30 min. Excess stain was removed by rinsing with distilled H_2O (3 x 100 µl). The wells were then air-dried and the remaining crystal violet extracted with 100 µl acidified methanol. Absorbance at 620 nm was determined using a Titertek Multiscan spectrophotometer.

Proliferation assays

The effects of synthetic peptides and growth factors on the growth of breast cancer and endothelial cells were determined as detailed.

Exponentially growing cells were harvested by trypsinisation, as previously described. After rinsing and resuspending in the relevant culture media (containing 10% FCS), the cells (100 µl aliquots) were dispensed into 96-well microtitre plates at a population density of 2 x 10^4 cells/well (6 wells per experimental condition). Cells were the incubated for 24 h at 37°C after which the media was removed and the wells rinsed with CFS (3 x 100 µl), to rid the plates of cells in suspension. Media was then replaced with that containing the relevant controls or treatment

supplements as detailed in individual experiments.

Cell numbers were evaluated spectrophotometrically at 620 nm, over the period of assay, after fixing with 10% formaldehyde and staining with crystal violet.

Proliferative responses were analysed using the Wilcoxan Rank test and significant differences at the $p < 0.05$ level, defined. Results of all growth studies were confirmed in at least 3 individual experiments.

Laminin attachment assay

Non-tissue culture grade 96-well plates, coated with 2.5 µg murine laminin in 50 µl CFS per well, were air-dried overnight at room temperature. Preliminary experiments indicated that cell attachment was concentration dependent; maximal binding occurred at a laminin coating of 2.5 µg/well. After rinsing with CFS (100 µl), the plastic was saturated with casein (0.2% in CFS). Plates were incubated at room temperature for 45 min then washed extensively with CFS (3 x 100 µl).

After removal of culture media, cells were detached from monolayers by the action of EGTA (0.02% in CFS) at 37°C. The cells were then centrifuged at 800 g for 2 min and the pellet resuspended in DMEM (T-47D) or RPMI (SK HEP-1).

Cells, at a population density of 10^6 cells/ml, were then aliquoted (1 ml) into microfuge tubes containing the individual peptide sequences and incubated for 1 h at 37°C. The cells (100 µl aliquots) were then added to the pre-coated multi-well plates and incubated for a further 60 min. Incubation media were removed and the wells washed with CFS (3 x 100 µl) to rid the plates of

non-adherent cells.

Attached cell numbers were evaluated spectrophotometrically at 620 nm after fixing with 10% formaldehyde and staining with crystal violet.

Attachment to $mEGF_{(33-42)}$

That $mEGF_{(33-42)}$ bound to the 67kDa laminin receptor was demonstrated using a biotinylated derivative of the peptide (Acetyl-C-[*S*-Acm]-VIGYSGDR-C-[*S*-Acm]-K-[N^{ε}-biotin]-amide) and a modification of the above laminin attachment assay.

Briefly, 96-well plates were coated with 100 µl/well streptavidin (5 µg/ml in carbonate buffer pH 9.6) and following an overnight incubation at 37°C, wells were washed with CFS (3 x 100 µl) and the plastic blocked with casein (0.2% in CFS). The plates were then incubated at room temperature for 45 min and washed with CFS as previously detailed. Biotinylated $mEGF_{(33-42)}$ in CFS was then aliquoted into the wells (0.1 ml of 100 µM) and the plates incubated for 3 h at 37°C.

After a further block with 0.2% casein, the wells were washed with with CFS (3 x 100 µl aliquots). Plates were kept at 4°C and used within 2 h.

Cells were prepared as above and pre-incubated for 1 h at 37°C with serial dilutions of anti-laminin receptor polyclonal (see below) or anti-EGF (R1) receptor monoclonal antibodies. Subsequent procedures were as detailed for the laminin attachment assay.

Laminin receptor binding determinations

a. Radiolabelling of laminin

^{125}I-laminin was prepared using ^{125}I-labelled sodium iodide (Amersham, UK) and immobilised chloramine-T (Iodobeads; Pierce, Illinois). Prior to use, the beads were washed with 500 µl phosphate buffer (pH 6.5) to remove excess reagent from the support. These were then allowed to air dry and individual beads added to a solution of carrier free Na^{125}I, diluted with iodination buffer (phosphate buffer pH 7.4). The beads were allowed to equilibrate for 5 min.

Laminin (10 µg in 10 µl) was then diluted into the iodination buffer and the system incubated at 20°C for 15 min. The solution was then removed from the reaction vessel and excess Na^{125}I and unincorporated $^{125}I_2$ separated from the iodinated protein by gel filtration on a GF-5 exclusion column (Pierce, Illinois). Iodinated laminin fractions were recovered at a specific activity of approximately 1.2 mCi/mg protein (864 Ci/mmol).

b. Competition binding estimation

Near confluent cultures of T47-D or SK HEP-1 cells were removed from monolayer with 0.02% EGTA and passed through a G-25 syringe needle to produce single cell suspensions. Aliquots of each cell type (10^6 cells/ml) were dispensed into separate Ependorf tubes (1 ml each) and pelleted. The cells were then resuspended in 1 ml ice-cold RPMI (SK HEP-1) or DMEM (T47-D) containing 0.1% BSA and either laminin or synthetic peptide at the concentrations indicated. Iodinated laminin was then added to each cell suspension to give a final ^{125}I-

laminin concentration of 0.1 nM (approximately 50,000 cpm). These mixtures were incubated overnight at 4°C.

The tubes were then microfuged at 10,000 g and the supernatant removed. After washing the pellet with 500 µl CFS, the remaining radioactivity was determined using a gamma radiation counter. Non-specific binding was determined by incubating cells with a 1000-fold molar excess of unlabelled laminin. All estimations were carried out in triplicate.

IC_{50} (concentration of unlabelled peptide required to produce 50% inhibition of radioligand binding binding) and EC_{50} (effective concentration for 50% inhibition of cell attachment) values were calculated using the Grafit curve-fitting programme (Erithacus Software, London, UK).

Migration assays

The method used was basically as described by Albrecht-Buehler (1977). Briefly, coverslips (22 x 22 mm) were treated in 5% detergent (7X; ICN Biomedicals) and washed in alcohol to remove grease. After drying, they were immersed in gelatin solution (Sigma, 300 Bloom; 0.5 g in 300 ml distilled H_2O) for 10 min. The coverslips were then dried by placing in a 70°C oven for 45 min.

Colloidal gold suspension was prepared by adding 11 ml distilled H_2O and 6 ml Na_2CO_3 (36.5 mM) to 1.8 ml $AuHCl_4$ (14.5 mM). The mixture was heated to 95°C at which point 1.8 ml of freshly prepared 0.1% formaldehyde solution was added; the temperature was maintained at 95°C. A suspension of colloidal gold was formed which was brown to absorbed light and blue to transmitted

light.

The gold suspension, was then added to petri dishes containing individual coverslips and the plates incubated at 37°C for 45 min. After washing with CFS (3 x 4 ml) to remove unattached gold particles, the coverslips were transferred to 6-well cluster dishes and UV sterilised.

Endothelial cells (SK HEP-1 and BRCE) in culture media (0.3 ml) were seeded onto the coverslips at an approximate density of 5 x 10^3 cells per well. The cells were allowed to plate down for 2 h at 37°C after which the treatments were added. Assay systems were maintained for a further 18 h after which the cells were fixed using 3% gluteraldehyde in cacodylate buffer (pH 7.2).

The assays were examined using a Leica DM1RB phase contrast microscope and Q500MC image analysis system incorporating a JVC TK-1280E colour camera (Leica, Milton Keynes, UK). The track images of at least 30 cells were video-captured and the area (representing migration response) determined for each. Statistical analysis of these areas was then carried out using Macintosh Instat software to perform both Kruskal-Wallis analysis of variance and Mann-Whitney *U*-tests in order to compare the treatment groups with controls.

RESULTS

Proliferative response

All peptides were examined for their ability to influence the growth of T47-D and SK-Hep 1 cell lines. At concentrations of peptide up to 100μM, no

significant effects were observed in either cell line.

Mechanism of action

We had shown in earlier studies that $mEGF_{(33-42)}$ could inhibit the EGF-stimulated angiogenic response in the early chick as well as blocking the basal and EGF-stimulated motility of primary and established endothelial cells.

During the present study we have shown that $mEGF_{(33-42)}$ also inhibits the angiogenic effects of laminin (Nelson et al 1995). Furthermore, we have demonstrated that the anti-angiogenic effects of $mEGF_{(33-42)}$ are mediated solely through the high affinity 67 kDa laminin receptor (67-LR) and not through the EGF receptor.

We have also confirmed that $mEGF_{(33-42)}$, $Lam.B1_{(925-933)}$ and laminin are equipotent in ^{125}I-laminin displacement receptor assays, and that both of the small peptidal ligands have similar potencies in specific laminin cell attachment assays.

In addition, we have shown that the commonly used chick angiogenesis models are not appropriate to the study of laminin mediated human angiogenesis: although we confirmed that $Lam.B1_{(925-933)}$ acts as a partial laminin antagonist in chick, we found it to be a pure agonist in mammalian cell lines. This is a highly significant point given that pharmaceutical companies (such as Angiotech, Vancouver, BC) are using the chick CAM assay as the sole screening method for the discovery of anti-angiogenic lead compounds. This may be inappropriate for use in human disease.

We were the first to show that the YIGSR-receptor is,

in fact, the 67 kda high affinity laminin receptor (67-LR). In collaboration with Professor Archer's team at the Department of Ophthalmology, Royal Victoria Hospital, Belfast, we have now determined that the 67-LR is preferentially expressed in new vessels during oxygen-induced retinopathy in neonatal mice (Nelson et al 1997a; manuscript in preparation).

Peptide antagonist development

The *N*-terminus of Lam.B1$_{(925-933)}$ is not necessary for receptor recognition and the agonist activity of YIGSR peptide (Ostheimer et al 1992, Kawasaki et al 1994).

However, alanine scanning of the starting peptide (mEGF$_{(33-42)}$) indicated that residues at positions 1, 2, 3, and 6 (peptidesVI, VII, VIII and X respectively), are essential for receptor mediated activities as determined by ^{125}I-laminin displacement and cell attachment to laminin through the 67-LR. Substitution of these individual residues by alanine leads to a dramatic decrease in receptor affinity observed as an increased IC_{50} (Table 2) and a parallel decrease in their ability to block adhesion to laminin (increased EC_{50}; Table 2). Characterisation of these analogues with regard to effects on motility, largely confirmed these findings although there was one exception; peptide VIII. Results from the migration assay identified this sequence (alanine for cysteine (P1)) as being a weak laminin agonist despite there being a much reduced response in the other two assays. It is suggested that this peptide may influence laminin receptor mediated migration through an alternative mechanism (Scott 1997).

Substitution at P10 (alanine for cysteine (peptide X)

retains both receptor binding and adhesion displacing activities but has the effect of changing the antagonistic parent into an agonist analogue. This reflects the response the agonism of $Lam.B1_{(925-933)}$, which also lacks the *C*-terminal cysteine, and suggests that this cysteine is not essential for receptor recognition, but is required for antagonism of $mEGF_{(33-42)}$.

Studies have reported that the positive charge offered by arginine (P9) is essential for the biological activity of $Lam.B1_{(925-933)}$ (McKelvey et al 1991, Kawasaki et al 1994). Glutamate substitution for arginine generates a negative charge at this position with corresponding loss of biological activities (Kawasaki et al 1994).

However, the substitution of arginine (P9) with positively-charged lysine (McKelvey et al 1991) also results in complete loss of ligand binding and biological activities, suggesting that the mere presence of a positive charge at this position is, in itself, insufficient for receptor recognition. Our modelling studies suggest that H-bonding of the guanidino group of the arginyl residue to the aromatic sidechain of the tyrosyl residue (P5) in the consensus sequence GY*X*G*X*R presents an acceptable motif for 67-LR activation by both $mEGF_{(33-42)}$ and $Lam.B1_{(925-933)}$.

Substitution of tyrosine (P5) with a conformationally restricted mimetic (tetrahydroisoquinoline-3-carboxylic acid; Tic-OH) in peptide V converted the antagonist $mEGF_{(33-42)}$ into an agonist. This residue substitution generates a predicted conformation unlikely to be able to form H-bonds. Although both receptor binding and adhesion responses were retained in this peptide the

loss of antagonism would suggest that H-bonding between tyrosine (P5) and the arginine (P9) is important for these antagonist activities.

Modelling studies suggested that citrulline (an uncharged arginine mimetic) would also be capable of forming this H-bonded motif.

It was found that replacement of arginine (P9) with citrulline (peptide IV) increased both receptor binding and inhibition of attachment to laminin substrata whilst retaining antagonist migratory response (Table 2), reinforcing our observation that it is not the positive charge that is required rather than a active conformation generated by hydrogen bonding. These findings thus identify H-bonding between P5 and P9 as being more important than the charge at the P9 arginine in determining antagonist activity.

Subsequent strategies involved the substitution of variant residues in the antagonistic $mEGF_{(33-42)}$ with those present in the agonistic $Lam.B1_{(925-933)}$ sequence (peptides I-III), in an effort to identify key amino acids in the *C*-terminal regions (P5-10) of the two ligands responsible for their contrasting bioactivities.

Substitution of isoleucine (P6) for serine (peptide I) resulted in both reduced receptor affinity and potency in displacement of cell adhesion to to laminin. However, this analogue retained weak antagonist activities in the motility assay. It is therefore of interest that studies on the YIGSR sequence indicate that residue substitution, at the position taken by isoleucine in the pentapeptide, are well tolerated and may increase potency (Kawasaki et al 1994).

Replacement of aspartate (P8) with serine (peptide II) resulted in a complete loss of biological function. as did peptide III encompassing both the former (isoleucine (P6) for serine) and latter (serine (P8) for aspartate) substitutions. Since this $mEGF_{(33-42)}$ analogue sequence (peptide II) encompasses the active YIGSR amino acid sequence agonist, it is suggested that this loss of activity may be attributed to the valine (P2) and isoleucine (P3) residues in the *N*-terminal half of $mEGF_{(33-42)}$. Alternatively, addition of a *C*-terminal cysteine to the YIGSR sequence is known to reduce potency (Kawasaki et al 1994). Additional peptides incorporating the valine (P2) and isoleucine (P3) substitutions are currently under investigation.

The determination of the minimum core peptide structure is ongoing and involves similar characterisation studies on a number of sequences truncated at the *C*-terminal.

These studies have thus identified an important antagonist of 67-LR mediated activies in peptide IV. The sequence, (AcC(Acm)-VIGYSGD-[Cit]-C-(Acm)-NH_2.), may provide an important template for anti-angiogenic drugs in that it is resistant to cleavage by trypsin-like proteases and has been identified as being more potent than $mEGF_{(33-42)}$ in our screening procedures.